

Thaw and Culture Details

| Cell Line Name | PENN001i-87-2 |
|-------------------------------------|---|
| WiCell Lot Number | DB36483 |
| Provider | University of Pennsylvania – Dr. Daniel Rader |
| Banked By | Penn Institute for Regenerative Medicine iPS Core Facility |
| Thaw and Culture Recommendations | WiCell recommends thawing 1 vial into 1 well of a 6 well plate. WiCell recommends thawing using ROCK Inhibitor for best results. |
| Culture Platform | Feeder Dependent |
| | Medium: hESC Medium (KOSR) |
| | Matrix: MEF |
| Protocol | WiCell Feeder Dependent Protocol |
| Passage Number | p13 These cells were cultured for 13 passages prior to freeze and post colony picking. Therefore, plated cells at thaw should be labeled passage 14. |
| Date Vialed | 20-July-2015 |
| Vial Label | iPS-87-098 SEV2 P13 7/20/2015 ZL |
| Biosafety and Use Information | Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans. |

Testing Performed by WiCell

| Test Description | Test Provider | Test Method | Test Specification | Result |
|------------------------|-------------------------|-----------------|------------------------------|--------|
| Post-Thaw Viable Cell | WiCell | SOP-CH-305 | Recoverable attachment after | Pass |
| Recovery | | | passage | |
| Identity by STR | UW Translational | PowerPlex 16 HS | Defines profile | Pass |
| | Research Initiatives in | System by | | |
| | Pathology Laboratory | Promega | | |
| Sterility | Biotest Laboratories | ST/07 | Negative | Pass |
| Mycoplasma | WiCell | SOP-QU-004 | Negative | Pass |
| Karyotype by G-banding | WiCell | SOP-CH-003 | Report karyotype | Pass |

Testing Reported by Provider

The Provider stated that some or all of the additional analyses listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

- SNP microarray
- Flow Cytometry (Tra1-60 and SSEA-4)
- Differentiation into hepatocytes
- Infinium[®] Expanded Multi-Ethnic Genotyping Array (MEGA^{EX})

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| Approval Date | Quality Assurance Approval |
|---------------|---|
| 23-June-2016 | 2/5/2018 X RK Quality Assurance Signed by: Kremers, Erik |

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The material provided under this certificate has been subjected to the tests specified and the results and data described herein are accurate based on WiCell's reasonable knowledge and belief. Appropriate Biosafety Level practices and universal precautions should always be used with this material. For clarity, the foregoing is governed solely by WiCell's Terms and Conditions of Service, which can be found at http://www.wicell.org/privacyandterms.



Short Tandem Repeat Analysis

Department of Pathology and Laboratory Medicine TRIP Laboratory (Molecular) http://www.pathology.wisc.edu/research/trip

Sample Report: 11771-STR Sample Name on Tube: 11771-STR 44.0 ng/μL, (A260/280=1.97) Sample Type: Cells Cell Count: ~2 million cells

Requestor: WiCell Research Institute Quality Department WiCell® info@wicell.org (888) 204-1782

Sample Date: N/A Receive Date: 08/22/16 Assay Date: 08/23/16 File Name: 160825 str jam Report Date: 08/26/16

| STR Locus | STR Genotype Repeat # | STR Genotype | | |
|------------|---|------------------------------|--|--|
| FGA | 16–18,18.2,19,19.2,20,20.2,21,21.2,22, 22.2, 23, 23.2, 24, 24.2, 25, 25.2, 26–30, 31.2, 43.2, 44.2,45.2, 46.2 | Identifying information has | | |
| ТРОХ | 6-13 | been redacted to | | |
| D8S1179 | 7-18 | protect donor | | |
| vWA | 10-22 | confidentiality. If | | |
| Amelogenin | X,Y | more information | | |
| Penta_D | 2.2, 3.2, 5, 7-17 | is required, please, contact | | |
| CSF1PO | 6-15 | WiCell's Technical | | |
| D16S539 | 5, 8-15 | Support. | | |
| D7S820 | 6-14 | | | |
| D13S317 | 7-15 | | | |
| D5S818 | 7-16 | | | |
| Penta_E | 5-24 | | | |
| D18S51 | 8-10, 10.2, 11-13, 13.2, 14-27 | | | |
| D21S11 | 24,24.2,25,25.2,26-28,28.2,29,29.2, 30, 30.2,31, 31.2,32,32.2,33,33.2, 34,34.2,35,35.2,36-38 | | | |
| TH01 | 4-9,9.3,10-11,13.3 | | | |
| D3S1358 | 12-20 | | | |

<u>Results:</u> Based on the 11771-STR cells submitted by WiCell QA dated and received 08/22/16, this sample (Label on Tube: 11771-STR) defines the STR profile of the human stem cell line PENN001i-87-2 comprising 26 allelic polymorphisms across the 15 STR loci analyzed.

<u>Interpretation:</u> No STR polymorphisms other than those corresponding to the human PENN001i-87-2 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggests that the 11771-STR sample submitted corresponds to the PENN001i-87-2 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~2-5%.

¹For this sample a microvariant exists at the D21S11 loci with a size between 33 and 33.2.

| X <i>RMB</i> Digitally Signed on 08/29/16 | X WMR Digitally Signed on 08/29/16 |
|---|---|
| TRIP Laboratory, Molecular | , PhD, Director / Co-Director UWHC Molecular Diagnostics Laboratory / UWSMPH TRIP Laboratory |

Testing was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only. Acknowledge TRIP in your publications, posters & presentations. For details, see: http://www.pathology.wisc.edu/research/trip/acknowledging TRIP agrees to maintain the confidentiality of any information provided to it in connection with its performance of this STR analysis on the same conditions as set forth in paragraph 2 of WiCell's Terms and Conditions of Service (http://www.wicell.org/media.acux/1a429b84-2b54-44a4-8ad8-5c05db93dd8a).

Sterility Report

| WiCell Research Institute, WiCell Quality Assurance | Inc. | | BIOTEST SAMPLE # VALIDATION # | 16080730 NG | | | |
|--|---|---------------------------|--|--|--|--|--|
| | | | TEST PURPOSE | NG | | | |
| PRODUCT | WA09-RB40917 11779, W STAN002i-161-1-DB31139 DB35052 11784, PENN066 PENN134i-61-26-DB35028 | 11782, PEN 6i-427-6-DB | NOO1i-87-2-DB36483 1 35047 11785, PENN074 | 1783, PENN002i-442-1- | | | |
| PRODUCT LOT | NA | | | | | | |
| STERILE LOT | NA | | BILOT | NA | | | |
| STERILIZATION LOT | NA | | BI EXPIRATION DATE | NA | | | |
| STERILIZATION DATE | NA | | DATE RECEIVED | 2016-08-11 | | | |
| STERILIZATION METHOD | EO | | TEST INITIATED | 2016-08-11 | | | |
| SAMPLING BLDG / ROOM | NA | | TEST COMPLETED | 2016-08-25 | | | |
| REFERENCE | Processed according to LAB-003: Sterility Test Procedure | | | | | | |
| | | | | 0 mL FTG. The sample was as monitored for a minimum | | | |
| | USP BI Manufacturers Speci Other | fications | | | | | |
| RESULTS Sterile | # POSITIVES # ⁻ 0 | TESTED 10 | POSITIVE CONTR NA | OL NEGATIVE CONTROL 2 Negatives | | | |
| COMMENTS NA | | _ | | | | | |
| REVIEWED BY | 3 | | DATE | 2040416 | | | |
| | | | | | | | |

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. Liability is limited to the costs of the tests. The uncertainty of measurement associated with the measurement result reported in this certificate is available from the organization upon request.

Biotest Laboratories # 9303 West Broadway Ave. # Brooklyn Park, MN 55445 # USA # (763) 315-1200

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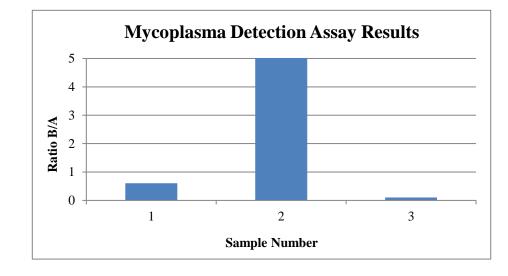
≋ STERIS



Mycoplasma Detection Assay Report Testing Performed by WiCell

Testing Performed by WiCell Lot Release Test August 5th, 2016 FORM SOP-QU-004.01 Version F Edition 01 Reported by: SM Reviewed by: JB Berthold Flash n' Glo 539

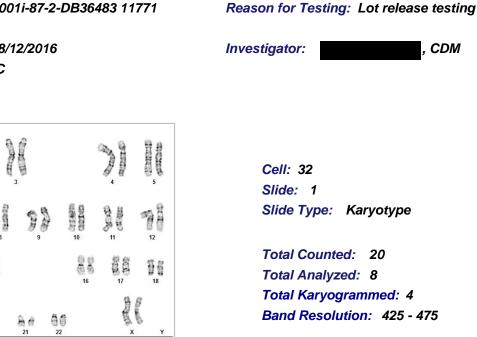
| | | Read | ing A | Α | Read | ling B | В | Ratio | | |
|---|-----------------------------|------|-------|-------|------|--------|------|-------|----------|-----------------------------|
| # | Sample Name | RLU1 | RLU2 | Ave | RLU1 | RLU2 | Ave | B/A | Result | Comments/Suggestions |
| 1 | PENN001i-87-2-DB36483 11771 | 214 | 211 | 212.5 | 127 | 129 | 128 | 0.60 | Negative | |
| 2 | Positive (+) Control | 373 | 381 | 377 | 8464 | 8514 | 8489 | 22.52 | Positive | |
| 3 | Negative (-) Control | 237 | 244 | 240.5 | 26 | 24 | 25 | 0.10 | Negative | |





Cell Line Gender: Female

Date Reported: Friday, August 19, 2016 Cell Line: PENN001i-87-2-DB36483 11771 Passage#: 14 Date of Sample: 8/12/2016 Specimen: iPSC Results: 46,XX



Interpretation:

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This is a normal karyotype. No clonal abnormalities were detected at the stated band level of resolution.

| Completed by: Reviewed and Interpreted by: A signed copy of this report is ava | | (ASCP) , PhD, FACMG quest. | |
|--|----------|----------------------------------|---------------|
| Date: | Sent By: | Sent To: | QC Review By: |

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results may not be relied upon by any other party without the prior written consent of the Director of the WiCell Cytogenetics Laboratory. The results of this assay are for research use only. If the results of this assay are to be used for any other purpose, contact the Director of the WiCell Cytogenetics Laboratory.

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